

Quantitative trait loci mapping for stomatal traits in interspecific hybrids of *Eucalyptus*

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Abstract. *Eucalyptus* is an important industrial species with tolerance to drought and salt stress. Genetic improvement activities including quantitative trait loci (QTL) mapping for pulping and adventitious rooting traits are in progress, but no information is available on the genomic regions on adaptive traits such as stomatal characteristics. In this study, an interspecific cross between *Eucalyptus tereticornis* and *E. grandis* was generated for the development of genetic map and QTL identification for stomatal traits. Simple sequence repeats (SSRs), inter-simple sequence repeats (ISSRs) and sequence related amplified polymorphism (SRAP) markers were used for genotyping the F_1 individuals. Parent-specific genetic maps (female, 1023.56 cM; male, 1049.64cM) and consensus map (1049.4 cM) were developed. QTL analysis was carried out to identify the chromosomal regions affecting stomatal density, area and pore length in adaxial and abaxial leaf surface. Seven QTLs were identified with phenotypic variation of 11.36 to 27.30% for stomatal traits when combined with growth and wood properties would have greater implications for generation of stress tolerant eucalypt hybrids with higher productivity and adaptability.

Keywords. linkage map; quantitative trait loci; stomata; stress tolerance; eucalyptus.

Introduction

Eucalyptus is grown in the tropics mostly for the pulpwood production. Globally, more than 21 Mha of *Eucalyptus* plantations are available (Midgley 2013) and produce 17.5% of world's paper pulp (Hart and Santos 2015). Next to Brazil, India occupies the second position in growing *Eucalyptus* with more than 4.4 Mha plantations (Xie 2015). The major species grown in India are *Eucalyptus camaldulensis* and *E. tereticornis*, due to their adaptability to drought and suitability for paper pulp production. *E.*

grandis is a subtropical species targeted for the breeding programmes worldwide because of its high pulp productivity. It is closely related to *E. tereticornis* and production of interspecific hybrids displaying hybrid vigour in terms of pulp yield and drought tolerance is possible (Madhibha *et al.* 2013). Despite, India being a major grower of *Eucalyptus*, studies on hybrid breeding and applied molecular genetics are very limited.

Physiological traits, important for plant growth responses and are often quantitative, suggesting that they are influenced by multiple genes. Patterning of stomata and their function in angiosperms are considered key determinants of growth rate and water balance in plants. This requires a better understanding of molecular mechanisms involved in stomatal distribution. Stomatal parameters like density and pore size have been reported to be correlated with the rate of photosynthesis and transpiration, drought, salinity, yield, elevated CO_2 in

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SM conducted marker polymorphism analysis in the laboratory, trait measurements, data analysis and drafted the manuscript. BVKW and DB participated in the seedling raising, field planting and maintenance of the mapping population. MGD participated in designing the experiments and finalized the manuscript. MA, NB, RD and SV participated in designing the experiments, conducted controlled pollination, seedling raising and field planting. YR conceived, organized and planned the research and finalized the manuscript.

woody and crop species (Gailing et al. 2008; Liu et al. 2017). Considerable progress has been made in detection of stomatal traits related quantitative trait loci (QTL) for salt and drought tolerance in crop species (Shahinnia et al. 2016; Liu et al. 2017). In Eucalyptus, genetic mapping and QTL analyses for various traits including growth and form, wood properties, adventitious rooting, flowering time, biotic and abiotic stresses and secondary metabolism have been reported (Henry and Kole 2014). Currently, only limited information is available on QTL governing stomatal characteristics and such OTL can pave way for the production of high yielding trees with abiotic stress tolerance, especially when combined with QTL for growth and wood properties. Hence, the objectives of the study were to develop a consensus linkage map for the interspecific cross E. tereticornis and E. grandis using simple sequence repeats (SSRs), intersimple sequence repeats (ISSRs) and sequence-related amplified polymorphism (SRAP) markers and to identify QTL associated with stomatal traits.

Material and methods

Plant materials

The mapping population used in this study consisted of the parents of the cross, *E. tereticornis* (Et86) × *E. grandis* (Eg9) and their F₁ individuals. Et86 is a selection from a seed production area (Pudukottai, 10°23'N, 78°51'E), while Eg9 is a selection from the provenance trail (13017, Lorne) located at Osamund (11°26'N, 76°37'E). Controlled pollination experiments were carried out during May 2010 and F₁ were field planted in 2011 at Panampally (10°47'N; 76°45'E), Kerala.

Marker genotyping

Two parents (Et86 and Eg9) and 98 F_1 individuals were genotyped with 114 SSR loci (table 1 in the electronic supplementary material at http://www.ias.ac.in/jgenet/), 13 ISSR primers (table 2 in the electronic supplementary material) and 13 SRAP primer combinations (table 3 in the electronic supplementary material). PCR amplifications and electrophoresis for SSR, ISSR and SRAP markers were carried out as reported by Arumugasundaram *et al.* (2011), Balasaravanan *et al.* (2005) and Li and Quiros (2001), respectively.

Generation of genetic linkage maps

ISSR and SRAP markers were named after the primer serial number and the approximate fragment size. Chi-square (χ^2) test was performed and markers that deviated

from the theoretical expected ratios were considered as distorted with a significance level (P < 0.01, P < 0.001and P < 0.0001) and used for analysis. Linkage analysis was performed using JoinMap 4.0 software (Van Ooijen 2011) with cross pollinator (CP) option. Linkage phase determination and grouping was done independently for the parental datasets. Linkage group (LG) was estimated by applying independence logarithm of odds (LOD) ranging from 1 to 10. SSR markers were assigned to the LG based on the information from published literature (Grattapaglia et al. 2015). Marker ordering was performed with regression mapping using the standard parameters and the markers that were mapped in the first and second round were included, maps were generated using Mapchart 2.1 software (Voorrips 2002). The parent-specific maps were integrated using the 'Combine groups for map integration' function of JoinMap to produce a consensus map using common SSR markers.

Stomatal morphology measurements and data analysis

Leaves of coppice shoots from the parents and hybrid individuals were selected for stomatal morphology studies. Stomatal observations were carried out in the 3rd and 4th fully expanded leaves on the 30th day after coppicing the plants, following the methodology described by James and Bell (2000). From each plant, two leaves were smeared with a thin film of nail varnish at the centre of the leaf blade on the abaxial and adaxial surface to make an imprint. Stomatal density per unit area (number of stomata per mm²) was determined at $400 \times$ magnification from 10 field views per slide using photomicroscope (Leitz, Japan). Images of the stomata were taken using an ultrascope fitted with a microscope and stomatal length (SL), width (SW) and pore length (PL) were measured using Scope Image 9.0. SL, SW and PL values were measured in micrometre (μ m) from abaxial and adaxial leaf surfaces.

Stomatal area (μm^2) was calculated using the following formula (Wang *et al.* 2012),

$$\frac{\mathrm{SL}\times\mathrm{SW}\times\pi}{4}$$

Descriptive statistics and one-way analysis of variance (ANOVA) were used to analyse the variation in stomatal traits between the parents and F_1 individuals using the SPSS ver. 22.0 software (IBM, Armonk, USA).

QTL analysis

QTL analysis was carried out using composite interval mapping (CIM) procedure in WinQTL Cartographer v2.5

Linkage	Total number of loci mapped Et86 Eg9		Common SSR markers	Total number of loci mapped in Et86 × Eg9	Size in cM	Mean distance	
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1	25	16	7	33	91.7	2.8	
2	21	12	6	27	99.8	4	
3	8	18	3	22	93.9	6.8	
4	13	28	7	33	83.1	2.5	
5	22	21	6	37	86.3	2.3	
6	15	9	2	21	97.5	4.6	
7	20	9	6	23	92.9	4	
8	31	9	7	33	92.3	2.8	
9	15	8	6	16	107.9	6.7	
10	22	28	3	47	114.7	2.4	
11	12	14	4	22	89.3	4.1	
Total	204	172	57	314	1049.4	_	
AVG	18.55	15.64	5.18	28.55	95.40	3.9	

Table 1. Mapping statistics for consensus map of *E. tereticornis* (Et86) \times *E. grandis* (Eg9).

(Wang *et al.* 2007) to identify QTLs for stomatal density, stomatal area and pore length in adaxial and abaxial leaf surfaces adopting backcross model. The LOD threshold was determined by permutation analysis with 1000 repetitions. The size of the analysis window was maintained at 10 cM with a walk speed (mapping resolution) of 1 cM and control marker number five. Stepwise regression (forward method) under model 6 and the LOD score corresponding to P = 0.05 was used to identify significant QTL.

Results

Marker production and linkage map generation

A total of 114, 115 and 129 SSR, ISSR and SRAP markers were generated, respectively and the details of SSR segregation pattern, ISSR and SRAP polymorphism status are given in tables 1, 2 and 3 in electronic supplementary material. These markers were used for framework map construction in female and male parents separately at LOD 3.0, where 11 groups, equivalent to the haploid chromosome number were formed. The details on marker types, segregation pattern, parent informativeness and chi-square significance are presented in table 4 in the electronic supplementary material. Details on number of markers mapped, segregation deviation (SD) markers for SSR, ISSR and SRAP of parental maps are given in tables 5 and 6 in the electronic supplementary material. The female map had 204 markers covering 1023.56 cM (table 5 and figure 1 in the electronic supplementary material). The male map had 172 markers with 1046.64 cM length (table 6 and figure 2 in the electronic supplementary material). The consensus map with a total length of 1049.4 cM had an average of 28.55 markers per LG, with an average spacing of 3.9 cM (table 1; figure 1).

Phenotypic variations in stomatal traits

The univariate statistics for stomatal density, stomatal area and pore length of the Et86×Eg9 mapping population is provided in table 2. Analysis of variance (*F*-test) indicated significant differences among the parents and F_1 hybrids for the traits such as stomatal density and area on the adaxial and abaxial surface of the leaf. However, pore length did not show significant difference for the abaxial surface. The absolute values of skewness and kurtosis were less than 1 for all traits, indicating that the phenotypic values of these traits were normally distributed and suitable for QTL analysis.

QTL analysis

Phenotypic variance (PV; R^2) controlled by these QTL ranged from 11.36 to 27.3% (table 3). No QTL were identified for the stomatal density on the abaxial leaf surface. One QTL of large effect (explaining 27% of phenotypic variance) in LG8 associated with marker locus M14E12-350 for stomatal pore length at the abaxial surface was identified. Two SSR loci (Embra204 in LG9 and Embra2 in LG11) were associated with the stomatal area at the abaxial surface and the PV explained were 12 and 14%, respectively. Two SRAP markers and three ISSR markers were associated with adaxial stomatal area, abaxial and adaxial stomatal pore length and stomatal density in adaxial surface of the leaves.



Figure 1. Consensus linkage map of the cross *Eucalyptus tereticornis* (Et86) and *E. grandis* (Eg9) showing QTL regions for stomatal traits. Distance among markers are indicated in cM to the left of the linkage groups; markers in red, blue and black colours are SSR, ISSR and SRAP, respectively and underlined markers are common between maternal and paternal maps. QTL for the traits analysed for depicted as coloured vertical bars to the right of the linkage groups 1 (stomatal area adaxial), 6 (stomatal density adaxial), 8 (pore length abaxial), 9 (stomatal area abaxial), 10 (pore length adaxial) and 11 (pore length abaxial).

	Density (r	no. mm^{-2})	Stomatal a	area (μ m ²)	Pore length (μ m)		
Genotype	Adaxial	Abaxial	Adaxial	Abaxial	Adaxial	Abaxial	
Et86	132.00 ± 22.9	213.00 ± 50.8	135.19 ± 26.6	92.26 ± 33.3	7.53 ± 1.2	5.22 ± 1.0	
Eg9	92.00 ± 14.7	168.00 ± 20.4	204.22 ± 34.0	133.60 ± 20.2	7.69 ± 1.6	6.17 ± 1.0	
F_1 individuals ($n = 98$)							
Mean	112.85	246.91	153.97	143.25	5.12	5.13	
Max.	169	351	244.11	238	9.17	8.85	
Min.	57	123	54.85	73.29	2.09	2.68	
Skewness	0.19	-0.21	0.32	0.64	0.4	0.21	
Kurtosis	-0.17	-0.45	0.74	0.33	0.6	-0.51	
SE (±)	22.43	49.37	32.81	32.27	1.32	1.28	
F-test	*	**	**	*	**	ns	

Table 2. Descriptive statistics and ANOVA for stomatal density, stomatal area and pore length of *E. tereticornis* (Et86), *E. grandis* (Eg9) and F_1 individuals.

**P < 0.01; *P < 0.05; ns, not significant; SE, standard error.

Table 3. QTLs for stomatal traits mapped in Et86 \times Eg9 using WinQTL Cartographer v2.5.

Traits	LG	Locus name	LOD	Left marker	Right marker	PV (%)	Additive
Stomatal density adaxial	6	ISSR3-550	4.15	ISSR22-425	M12E10-146	14.9	21.07
Stomatal area abaxial	9	EMBRA204	4.37	CD519	EMBRA1811	12.4	-23.21
Stomatal area abaxial	11	EMBRA2	3.93	M11E11-297	ISSR3-850	14.4	-24.81
Stomatal area adaxial	1	M2E9-292	3.42	EMBRA303	M1E9-388	11.4	22.85
Pore length abaxial	8	M14E12-350	11.34	ISSR27-1000	EMBRA1468	27.3	-1.69
Pore length abaxial	11	ISSR22-475	6.8	ISSR3-850	M12E13-194	14.8	1.39
Pore length adaxial	10	ISSR27-675	7.37	M11E11-200	ISSR11-900	24	-1.51

LG, linkage group; PV(%), phenotypic variation (R^2).

Discussion

Genetic linkage map

A number of linkage maps are reported in Eucalyptus including high-density maps in interspecific crosses of E. grandis and E. urophylla, the hybrid combinations predominantly grown in Brazil, Australia and South Africa (Freeman 2014). Only very few linkage maps are available for the hybrid combinations with E. tereticornis as parent species. The individual parent-specific map generated in this study for E. tereticornis (Et86) was 1023.56 cM. Yu et al. (2012) reported a map of size 1488 cM with 21 LGs using expressed sequence tag-based cleaved amplified polymorphic sequence markers (EST-CAPS) and random amplified polymorphic DNA (RAPD) markers in E. tereticornis. Recently, for the same species, Li et al. (2015) generated linkage map of 1241 cM length with 585 loci distributed in 11 LGs. In E. grandis, several genetic maps were developed using different types of DNA markers and the linkage map length ranged from 925 cM (Kullan et al. 2012) to 1216 cM (Gion et al. 2011). The present study also observed the map length of 1046.64 cM with 172 markers belonging to SSR, ISSR and SRAP. Most of the reports on Eucalyptus maps constructed till date reported the haploid number of chromosomes, despite of various types of DNA markers, pedigree structure and mapping software packages (Freeman 2014) and the present study also showed marker distribution on 11 LGs.

QTL mapping for stomatal traits

Size, density and pattern of leaf stomata play an important role in plant growth and yield. Genetic relationships between stomatal density and size with yield were documented in cereals (Shahinnia et al. 2016). In Quercus, based on the QTL for stomatal density, their role in adaption towards climate change was discussed (Gailing et al. 2008). The aim of this study was to identify the QTL regions controlling stomatal density, stomatal area and pore length on the adaxial and abaxial surface of the Et86 \times Eg9 mapping population. The marker loci M14E12-350 in LG8 showed cosegregation for stomatal pore length at the abaxial surface with PV of 27.3%. The higher PV (> 25%) recorded in this study could be attributed towards the use of smaller populations, lack of multienvironment measurements and wide physical linkage between adjacent causal loci within single crosses (Hall et al. 2016).

This result is a step forward in understanding the function of these loci and to correlate their role in yield and biotic and abiotic stress tolerance. The accuracy of QTL mapping depends on several factors such as density of genetic map, genetic architecture of the trait, phenotyping efforts, statistical methods and population size (Semagn *et al.* 2010). In the present study, the size of mapping population is small and established only in a single environment. Nevertheless, practical applications for such QTL-linked markers will require validating them in different genetic backgrounds and environments.

Stomatal parameters like density, area and pore length gain significance in Eucalyptus because of the variation expressed towards abiotic stress tolerance, photosynthetic and transpiration efficiency. These characteristics affect the growth and productivity of Eucalyptus, where stomata play a major role in the control of water evaporation, gas exchange and pathogen entry (Tong et al. 2016). Stomata size and density are under a complex genetic control and thus provide multiple levels of regulation for stomatal functions (Chaves et al. 2016). Therefore, a better understanding of the genetic control of stomatal parameters is an important aspect in breeding of Eucalyptus for productivity, abiotic stress tolerance and disease resistance (Héroult et al. 2013: Tong et al. 2016). Wider variations in stomatal density, area and pore length in Et86 \times Eg9 provide an opportunity to assess their influence on various physiologically adaptive characteristics of hybrid individuals. The individuals which compromise between high water use efficiency and leaf cooling capacity would be the best-adapted genotypes under semiarid conditions (Chaves et al. 2016). Thus, correlation of stomatal traits with yield and adaptability would help to improve the productivity of *Eucalyptus* plantations in arid conditions.

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References

- Arumugasundaram S., Ghosh M., Veerasamy S. and Ramasamy Y. 2011 Species discrimination, population structure and linkage disequilibrium in *Eucalyptus camaldulensis* and *Eucalyptus tereticornis* using SSR markers. *PLoS One.* 6, e28252.
- Balasaravanan T., Chezhian P., Kamalakannan R., Ghosh M., Yasodha R., Varghese M. and Gurumurthi K. 2005 Determination of inter- and intra-species genetic relationships among six *Eucalyptus* species based on inter-simple sequence repeats (ISSR). *Tree Physiol.* 25, 1295–302.

- Chaves M. M., Costa J. M., Zarrouk O., Pinheiro C., Lopes C. M. and Pereira J. S. 2016 Controlling stomatal aperture in semiarid regions-The dilemma of saving water or being cool? *Plant Sci.* **251**, 54–64.
- Freeman J. S. 2014 Molecular linkage maps of *Eucalyptus*: strategies, resources and achievements. In *Genetics, genomics and breeding of eucalyptus* (ed. R. J. Henry and C. Kole), pp. 58– 74. CRC Press, Boca Raton.
- Gailing O., Langenfeld-Hyeser R., Polle A. and Finkeldey R. 2008 Quantitative trait loci affecting stomatal density and growth in a *Quercus robur* progeny: implications for the adaptation to changing environments. *Glob. Change Biol.* 14, 1934–1946.
- Gion J. M., Carouche A., Deweer S., Bedon F., Pichavant F., Charpentier J. P. et al. 2011 Comprehensive genetic dissection of wood properties in a widely grown tropical tree: *Eucalyptus*. *BMC Genomics.* **12**, 301.
- Grattapaglia D., Mamani E. M. C., Silva-Junior O. B. and Faria D. 2015 A novel genome-wide microsatellite resource for species of *Eucalyptus* with linkage-to-physical correspondence on the reference genome sequence. *Mol. Ecol. Resour.* 15, 437–448.
- Hall D., Hallingbäck H. R. and Wu H. X. 2016 Estimation of number and size of QTL effects in forest tree traits. *Tree Genet. Genomes.* **12**, 110.
- Hart P. W. and Santos R. B. 2015 Changing the face of short fiber a review of the *Eucalyptus* revolution. *Tappi J.* 14, 353–359.
- Henry R. and Kole C. 2014 *Genetics, genomics and breeding of eucalypts.* Boca Raton, CRC Press.
- Héroult A., Lin Y. S., Bourne A., Medlyn B. E. and Ellsworth D. S. 2013 Optimal stomatal conductance in relation to photosynthesis in climatically contrasting *Eucalyptus* species under drought. *Plant Cell Environ.* 36, 262–274.
- James S. A. and Bell D. T. 2000 Leaf orientation, light interception and stomatal conductance of *Eucalyptus globulus* ssp. *globulus* leaves. *Tree Physiol.* **20**, 815–823.
- Kullan A. R. K., Van Dyk M. M., Jones N., Kanzler A., Bayley A. and Myburg A. A. 2012 High-density genetic linkage maps with over 2,400 sequence-anchored DArT markers for genetic dissection in an F2 pseudo-backcross of *Eucalyptus grandis* × *E. urophylla. Tree Genet. Genomes.* 8, 163–175.
- Li F., Zhou C., Weng Q., Li M., Yu X., Guo Y. et al. 2015 Comparative genomics analyses reveal extensive chromosome colinearity and novel quantitative trait loci in *Eucalyptus*. *PLoS One*. **10**, e0145144.
- Li G. and Quiros C. F. 2001 Sequence-related amplified polymorphism (SRAP), a new marker system based on a simple PCR reaction: its application to mapping and gene tagging in *Brassica. Theor. Appl. Genet.* **103**, 455–461.
- Liu X., Fan Y., Mak M., Babla M., Holford H., Wang F. et al. 2017 QTLs for stomatal and photosynthetic traits related to salinity tolerance in barley. *BMC Genomics.* 18, 9.
- Madhibha T., Murepa R., Musokonyi C. and Gapare W. 2013 Genetic parameter estimates for interspecific *Eucalyptus* hybrids and implications for hybrid breeding strategy. *New Forests.* **44**, 63–84.
- Midgley S. J. 2013 Making a difference: celebrating success in Asia. *Aust. For.* **76**, 73–75.
- Semagn K., Bjornstad A. and Xu Y. 2010 The genetic dissection of quantitative traits in crops. *Elec. J. Biotech.* (http://doi.org/10.2225/vol13-issue5-fulltext-21).
- Shahinnia F. F., Roy J. J. L., Laborde B. B., Sznajder B. B., Kalambettu P. P., Mahjourimajd S. S. et al. 2016 Genetic association of stomatal traits and yield in wheat grown in low rainfall environments. *BMC Plant Biol.* 6, 150.

- Tong Y. G., Ding X. X., Zhang K. C., Yang X. and Huang W. 2016 Effect of the gall wasp *Leptocybe invasa* on hydraulic architecture in *Eucalyptus camaldulensis* plants. *Front. Plant Sci.* 7, 130.
- Van Ooijen J. W. 2011 Multipoint maximum likelihood mapping in a full-sib family of an outbreeding species. *Gen. Res.* 93, 343–349.
- Voorrips R. 2002 MapChart: software for the graphical presentation of linkage maps and QTLs. J. Hered. 93, 77–78.
- Wang H., Shi H., Yang R., Liu J. and Yu Y. 2012 Stomatal characteristics of greening plant species in response to different urban atmospheric environments in Xi'an China. J. Food Agric. Environ. 10, 1524–1529.

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- Wang S., Basten C. J. and Zeng Z. B. 2007 Windows QTL cartographer 2.5. https://brcwebportal.cos.ncsu.edu/qtlcart/ WQTLCart.htm.
- Xie Y. 2015 Current situation and development of *Eucalyptus* research in China. In *Abstracts IUFRO eucalypt conference on scientific cultivation and green development to enhance the sustainability of eucalypt plantations.* Zhanjiang, Guangdong, China (www.iufro.org/download/file/22326/5738/20803-zhanjiang15-abstracts_pdf/).
- Yu X., Guo Y., Zhang X., Li F., Weng Q., Li M. et al 2012 Integration of EST-CAPS markers into genetic maps of *Eucalyptus urophylla* and *E. tereticornis* and their alignment with *E. grandis* genome sequence. *Silvae Genet.* **61**, 247– 255.